RT-PCT Test Quality Control

-Chen Yu Hui BGI Research

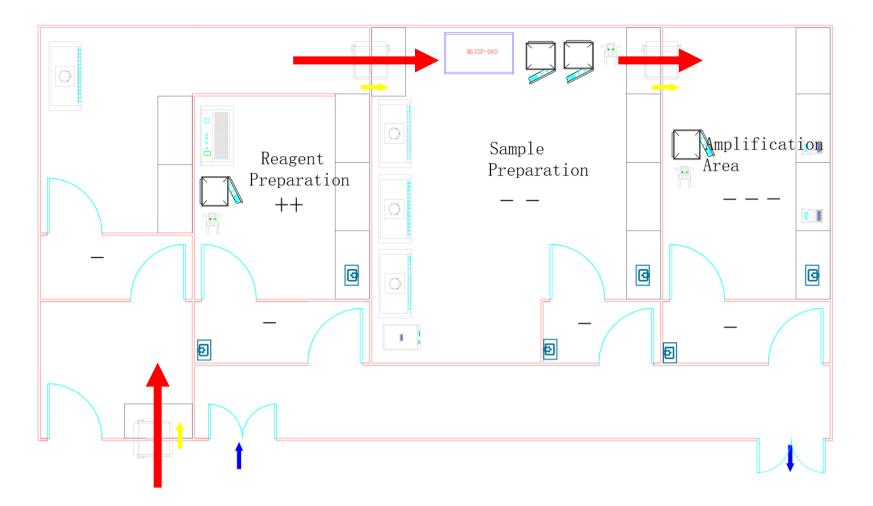
Prior to sample handling
Check points during experiment
Quality control of test data

Sample Collection——Sample Pretreatment——RNA Extraction——qPCR Reaction——Result Calling

Prior to sample handling

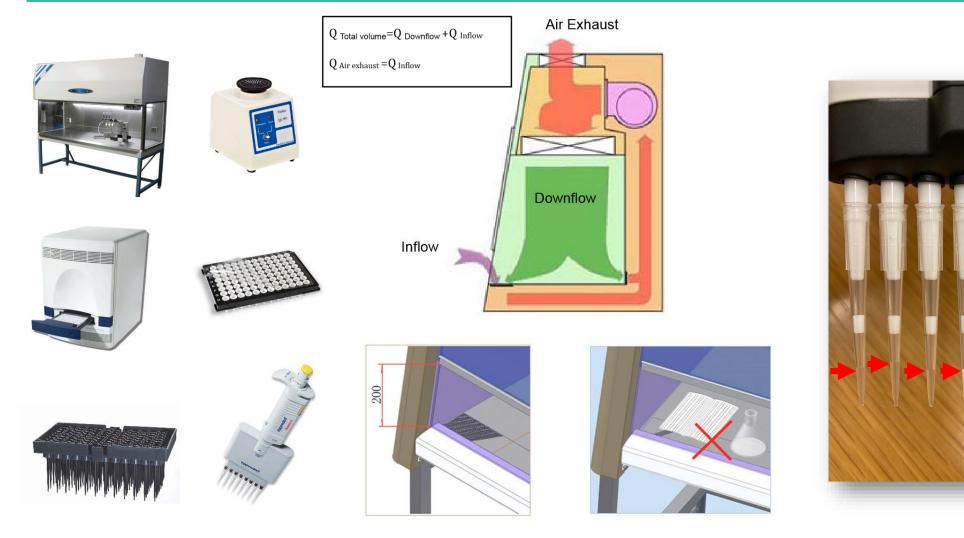
Laboratory layout and designated function of each area
 Equipment Maintenance
 Reagent and Consumables QC

Prior to sample handling - Laboratory layout



*Designated Function *Unidirectional Workflow *Proper pressure to contain nucleic acid in the expected area

Prior to sample handling - Maintenance; Consumables QC



#Function #Calibration #Proper Usage Training

Prior to sample handling – Reagent QC & Verification

Key reagent: Nucleic Acid Extraction Kit and RT-PCT Kit

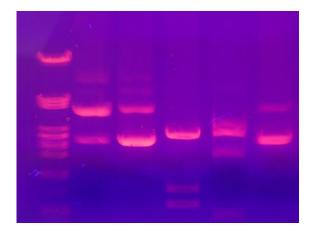
#Nucleic Acid Extraction Kit: Concentration and Purity

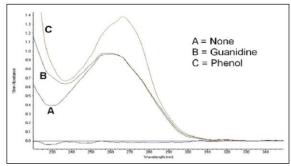
- Nanodrop or Qubit:A260/A280 & Electrophoresis
- Ct value: Control samples with known concentration

#RT-PCT Kit

- Sensitivity & Specificity
- Repeatability & Limit of Detection (LOD)

Conc.	Channel	1	2	3	4
100000000000000000000000000000000000000	FAM				
10000copies/ml	VIC				
5000.00mi00/ml	FAM				
5000copies/ml	VIC				
2500.00ming/ml	FAM				
2500copies/ml	VIC				



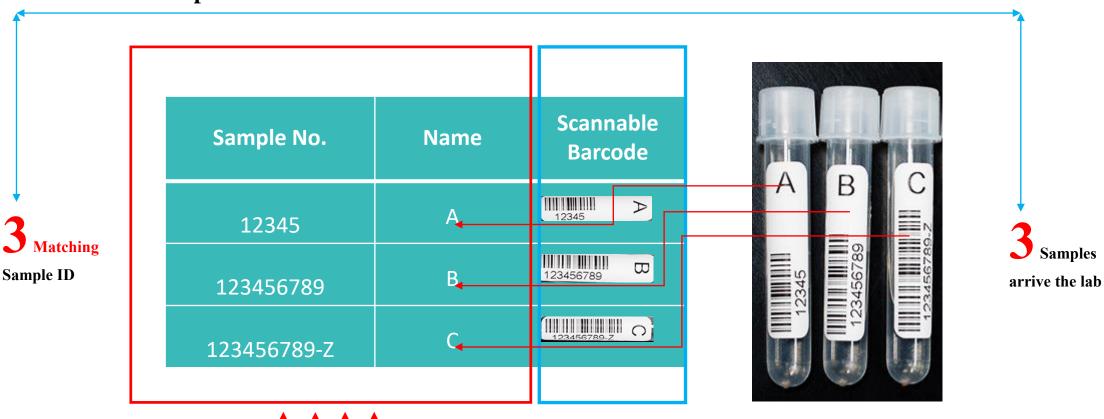


*It is suggested, at least, to have two types of RT-PCT kit for result verification

Check points during experiment

Sample arrival status & Sample registration
 Sample assignment into consumables
 RNA extraction & RT-PCR reaction

Check points - Sample arrival status & Sample registration



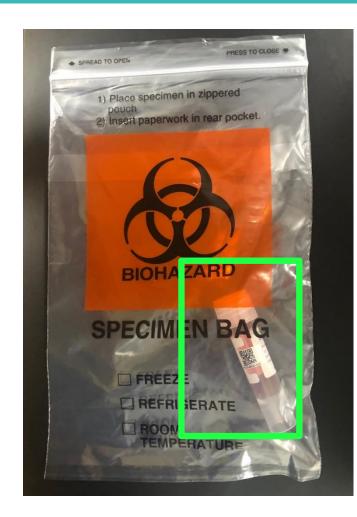
Samples should reach the lab in the box at DESIABLE TEMPERATURE

Provide Sample No. and Name of each sample in excel sheet to the lab BEFORE ship out the samples

HARD COPY of Information sheet with **SCANNABLE** barcode on the sheet is acceptable.

Sample Information/barcode provided in Hand-writing form will end up in sample rejection!!!

Check points-Sample arrival status & Sample registration







ONE TUBE, ONE BAG

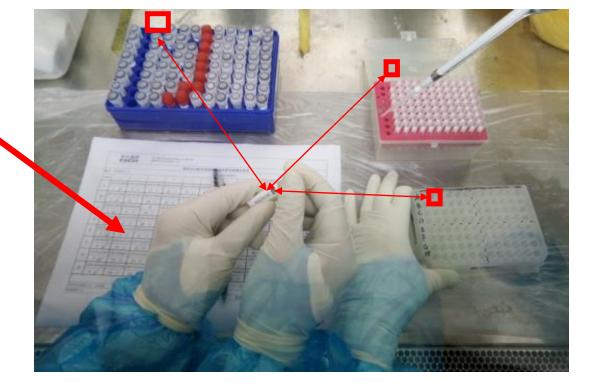
*Scannable barcdoe

*Barcode paper should be *heat-resistence*

One sample leaks, All samples will be rejected

Check points - Sample assignment

		COV	ID-19检	测任务	单/ COV	ID-19 \$	Sample I	Nucleic .	Acid Te	st Recoi	ď		A1	KA0004817
板号/Plate ID.:20200407-1 排单人Arrange Staff: HZB 排单时间/Time:							提取试剂配液编号Extraction Reagent Batch ID: 0318-E1						B1	KA0003576
sample ID	1	2	3	4	5	6	7	8	9	10	н	L.	C1	KA0004837
А	KA0004817	KA0003944	KA0004804	KA0004845									11	KA0004827
A	1 🛛	9 🗆	17 🗆	25 🗆	33 🛛	41 🛛	49 🛛	57 🛛	<mark>65</mark> □	73 🛛	81 🛛	89 🗆	E1	KA0004810
В	KA0003576	KA0003572	KA0004822	KA0004818									F1	KAD. 3578
D	2 🗆	10 🗌	19 🗆	26 🗆	34 🛛	42 🛛	50 🗆	58 🗆	<mark>66</mark> □	74 🛛	82 🛛	90 🗆	G1	KA0004833
с	KA0004837	KA0003585	Blank	KA0003591									H1	KA0003586
C	3 🛛	11 🛛	19 🗆	27 🛛	35 🗆	43 🛛	51 🛛	<u>59</u> 🛛	<mark>67</mark> □	75 🛛	83 🛛	91 🛛	A2	KA0003944
D	KA0004827	KA0003583	KR000+8+5	KA0003210									B2	KA0003572
ע	4 🛛	12 🛛	20 🗆	28 🗆	36 🛛	44 🛛	52 🗆	<u>60</u>	68 🛛	76 🛛	84 🛛	92 🗆	C2	KA0003585
Е	KA0004810	KA0003574	KA0004842										D2	KA0003583
E	5 🗆	13 🛛	21 🗆	29 🗆	37 🛛	45 🛛	53 🛛	<u>61</u>	<mark>69</mark> □	77 🛛	85 🛛	93 🛛	E2	KA0003574
F	KA0003578	KA0004844	KA0004806										F2	KA0004844
г	6 🗌	14 🛛	22 🗆	30 🗆	38 🗆	46 🛛	54 🛛	62 🛛	70 🗆	78 🛛	86 🛛	94 🛛	G2	KA0003597
G	KA0004833	KA0003597	KA0004840										H2	KA0003214
0	7 🛛	15 🛛	23 🗆	31 🗆	39 🗆	47 🛛	55 🗆	63 🛛	71 🛛	79 🛛	87 🛛	95 🗆	A3	KA0004804
Н	KA0003586	KA0003214	KA0004830									PC	B3	KA0004822
н	8 🗆	16 🛛	24 🗆	32 🗆	40 🗆	48 🗆	56 🗆	64 🛛	72 🗆	80 🗆	88 🗆	96 🛛	C3	Blank
备注/Remark: qPCRit济幅浓编号/qPCR Reagant Mix Batch ID: 0318-Q1									D3	KA0004843				
取祥操作人/Sampling Staff. 复核人/Verifier. 取样时间/Sampling Time:									E3	KA0004842				
提取操作人/Nucleic Acid Extraction Staff: 复核人/Verifier: 提取仪器编号/Nucleic Acid Extraction MGISP-960 No.: 6 ト机操作人 aPCR Operator: aPCR(公编号/aPCR Machine No.: 上机时间/aPCR Experiment Time:											F3 G3	KA0004806 KA0004840		

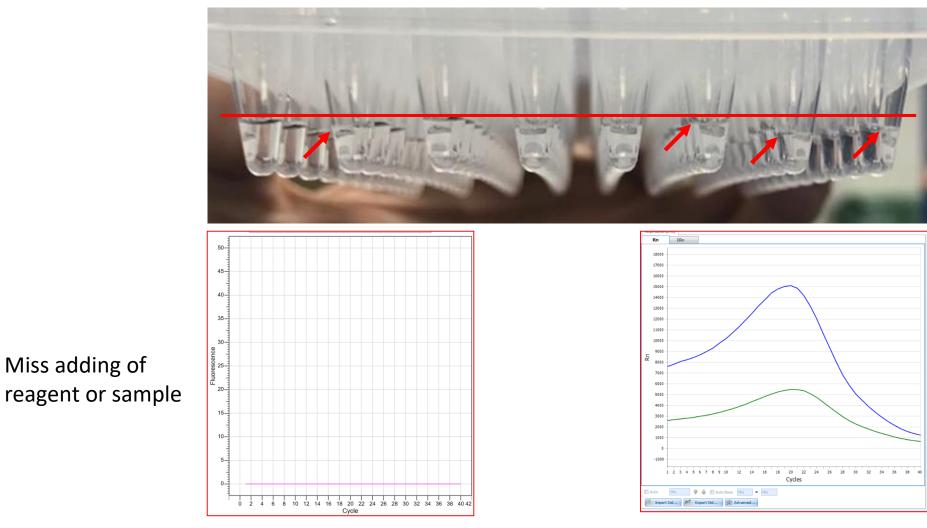


Sample layout for RNA extraction

Sample loading verified by two technicians

Check points - RNA extraction & RT-PCR reaction

Sample volume matters!



Evaporation

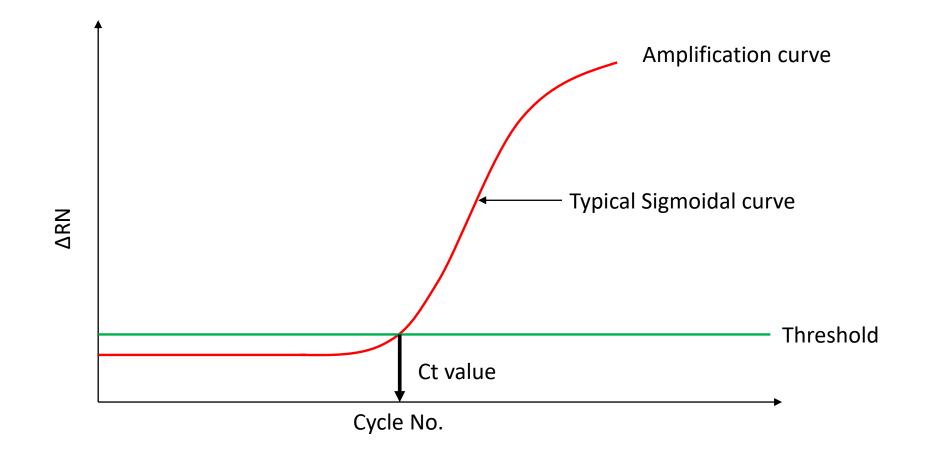
NO Signal Captured

Miss adding of

Unusual Amplification Plot

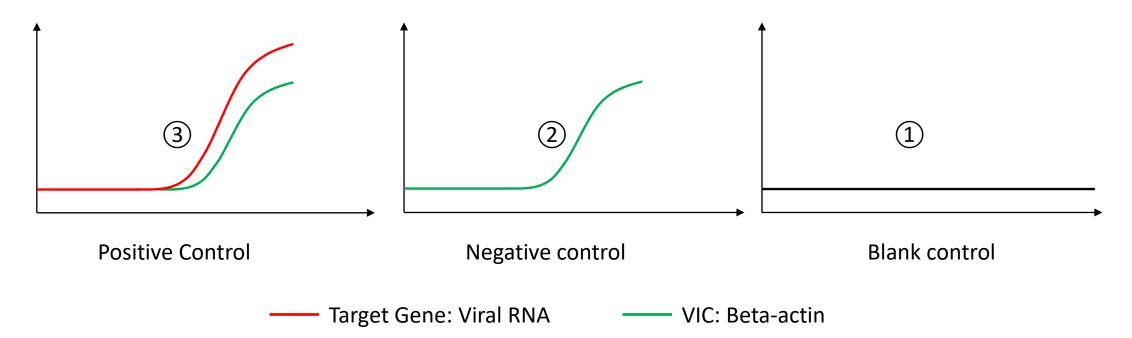
Quality control of test data - Key Concepts

- **Baseline**: The signal level during the initial cycles of PCR, usually cycles 3 to 15, in which there is little change in fluorescent signal.
- Threshold: A statistically significant increase over the calculated baseline signal. Usually sets the threshold at 10 times the standard deviation of the fluorescence value of the baseline.
- **Ct**: Cycle number at which the fluorescent signal of the reaction crosses the threshold. So no cross, no Ct.



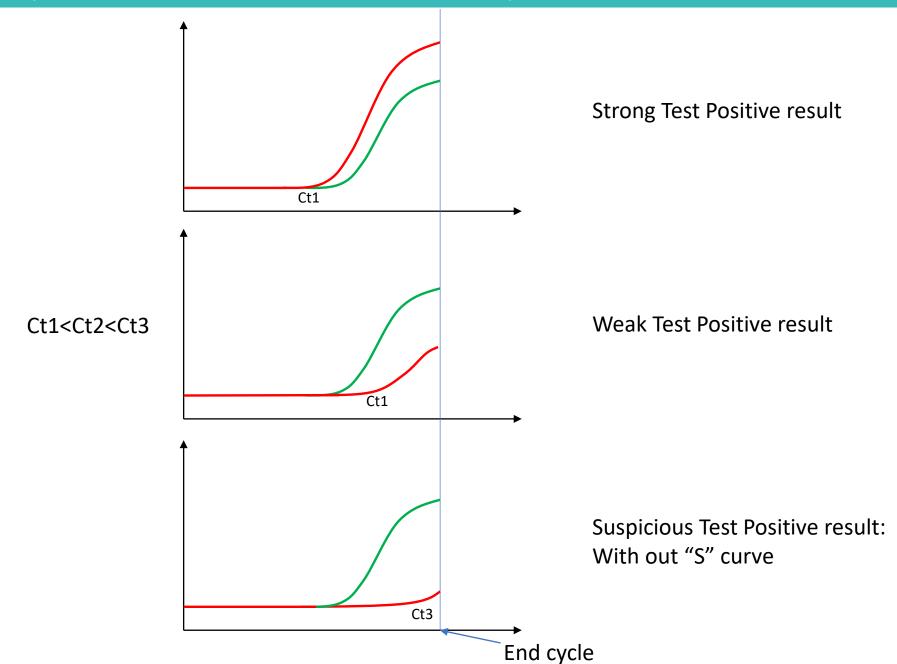
Quality control of test data – Control samples

- Positive Control: To check whether Viral RNA can be detected
- Blank control: To monitor the contamination level or background noise level
- Negative control: To check whether beta-actin gene can be amplified successfully without Viral RNA signal

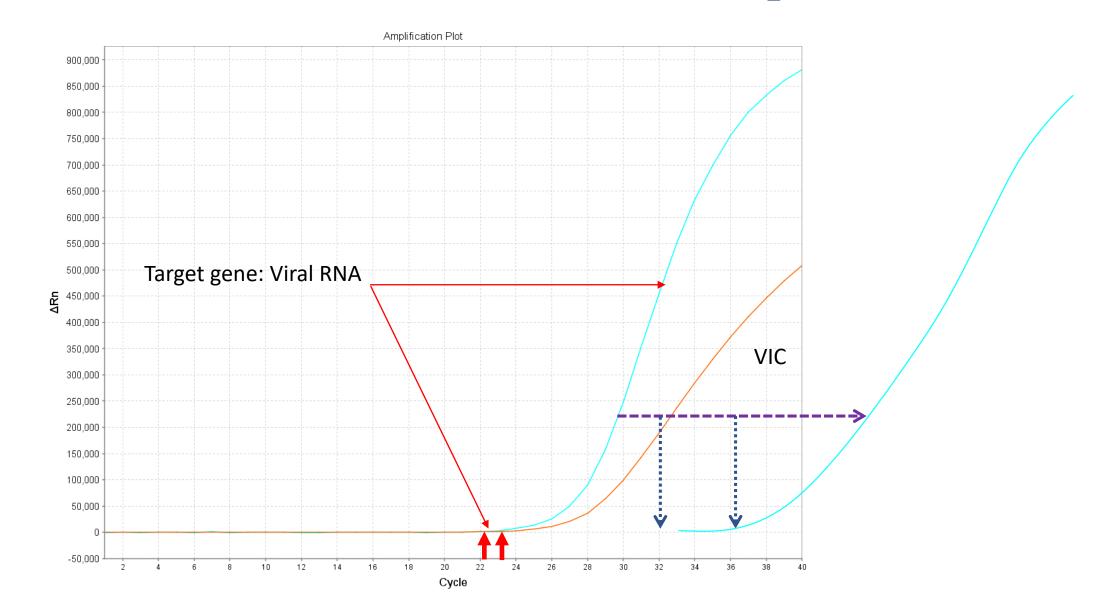


Both viral RNA and beta-actin DNA/RNA should be extracted and then amplified during qPCR reaction, which realizes the quality control of nucleic acid extraction. No beta-actin gene amplification or unusual amplification plot will lead to retest.

Quality control of test data – Example of result with Ct value



Quality control of test data – Example of result with Ct value

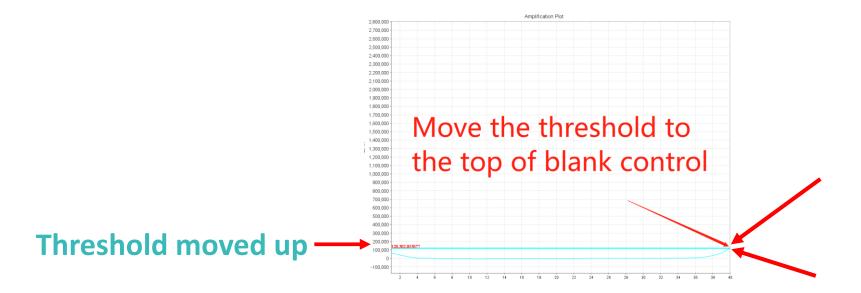


Why non-specific amplification in blank control?

- 1) Primer design is not perfect causing primer dimer formation leading to amplification in the end of PCR cycles;
- 2) Dye in the reagent kit partly degrades evidently in the end of cycles;
- 3) Primer in the kit is too concentrated, thus generating non-specific peaks for blank controls;
- 4) Slight contamination during sample mixing;
- 5) Lab environment is not clean enough with accumulated DNA/RNA aerosol;
- 6) 7500 is not well calibrated;

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- Always, *repeating the test and * thoroughly clean the laboratory area and related equipment will alleviate this situation. Separating RNA extraction and reagent mixing in different Bio-safety cabinets is suggested.
- Manually move the threshold to cover the noise and reanalyse Ct value for the rest of sample is suggested before data interpretation.



THANK YOU!