# **RT-PCT Test Quality Control**

-Chen Yu Hui BGI Research

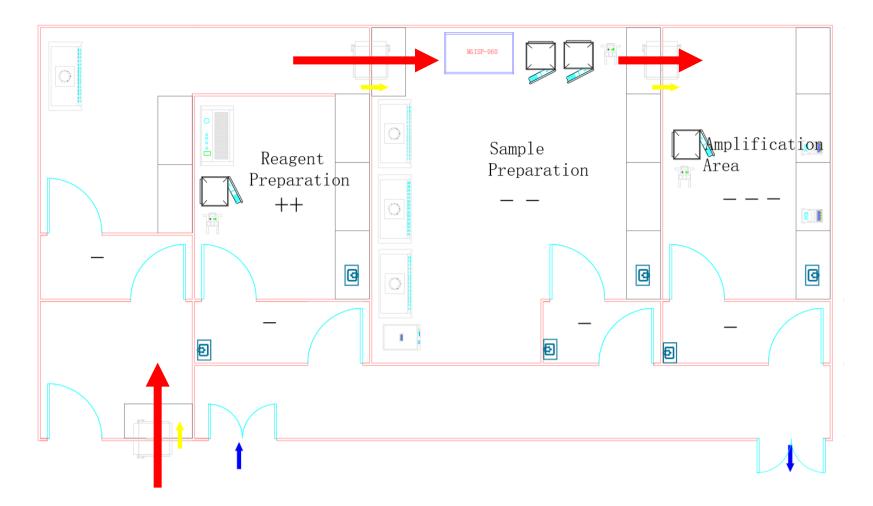
Prior to sample handling
Check points during experiment
Quality control of test data

Sample Collection——Sample Pretreatment——RNA Extraction——qPCR Reaction——Result Calling

### Prior to sample handling

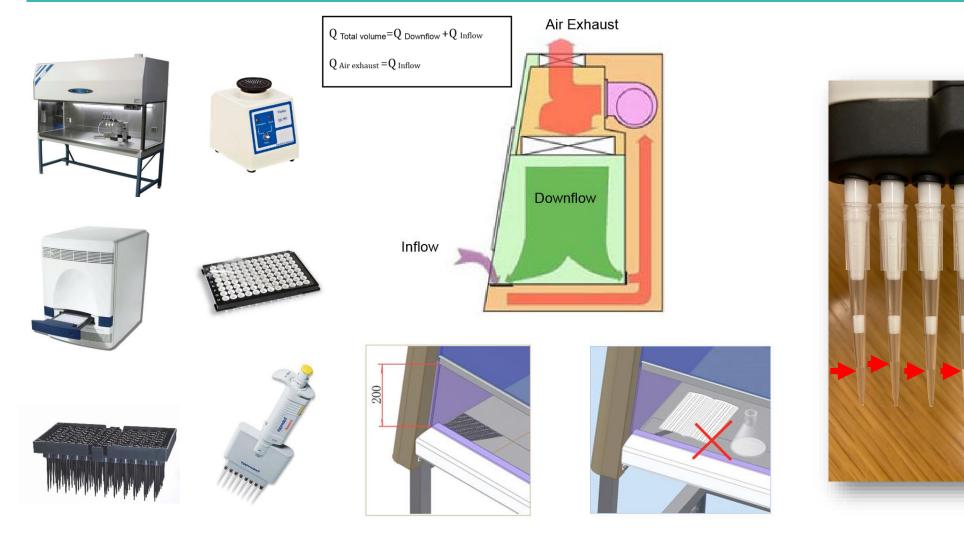
Laboratory layout and designated function of each area
 Equipment Maintenance
 Reagent and Consumables QC

# Prior to sample handling - Laboratory layout



\*Designated Function \*Unidirectional Workflow \*Proper pressure to contain nucleic acid in the expected area

# Prior to sample handling - Maintenance; Consumables QC



**#Function #Calibration #Proper Usage Training** 

# Prior to sample handling – Reagent QC & Verification

Key reagent: Nucleic Acid Extraction Kit and RT-PCT Kit

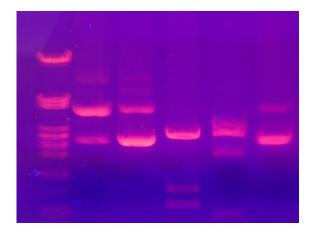
#### #Nucleic Acid Extraction Kit: Concentration and Purity

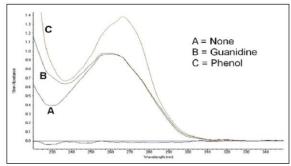
- Nanodrop or Qubit:A260/A280 & Electrophoresis
- Ct value: Control samples with known concentration

#### **#RT-PCT** Kit

- Sensitivity & Specificity
- Repeatability & Limit of Detection (LOD)

Conc.	Channel	1	2	3	4
100000000000000000000000000000000000000	FAM				
10000copies/ml	VIC				
5000.00mi00/ml	FAM				
5000copies/ml	VIC				
2500.00ming/ml	FAM				
2500copies/ml	VIC				



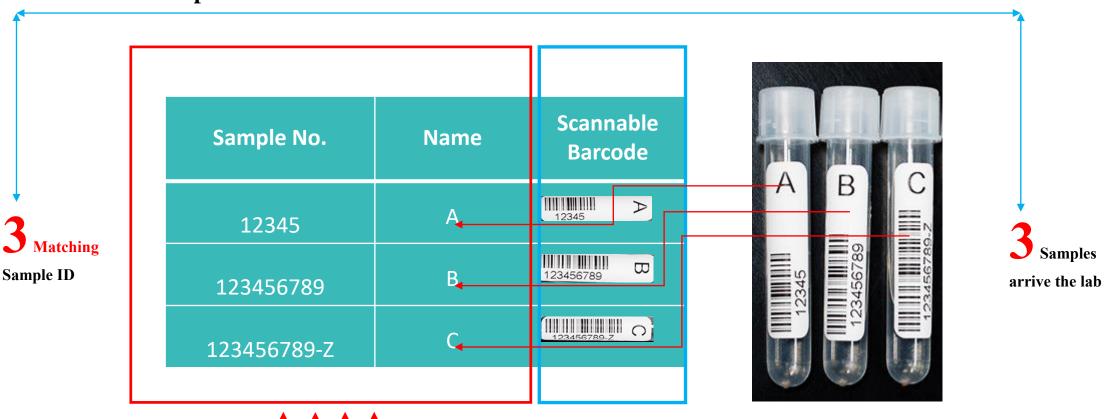


\*It is suggested, at least, to have two types of RT-PCT kit for result verification

# Check points during experiment

Sample arrival status & Sample registration
 Sample assignment into consumables
 RNA extraction & RT-PCR reaction

# Check points - Sample arrival status & Sample registration



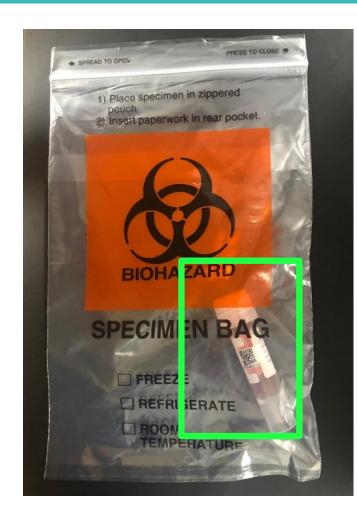
#### **Samples should reach the lab in the box at <b>DESIABLE TEMPERATURE**

Provide Sample No. and Name of each sample in excel sheet to the lab BEFORE ship out the samples

HARD COPY of Information sheet with **SCANNABLE** barcode on the sheet is acceptable.

Sample Information/barcode provided in Hand-writing form will end up in sample rejection!!!

# Check points-Sample arrival status & Sample registration







**ONE TUBE, ONE BAG** 

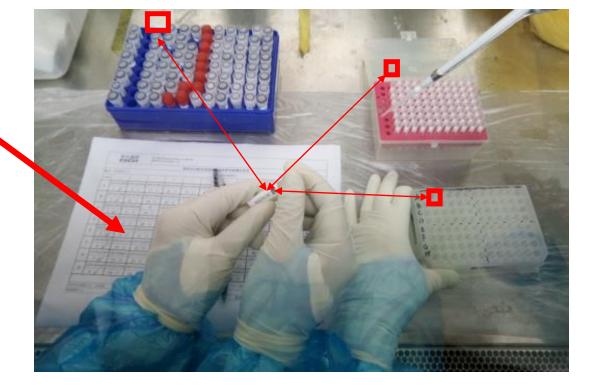
\*Scannable barcdoe

\*Barcode paper should be *heat-resistence* 

## One sample leaks, All samples will be rejected

# Check points - Sample assignment

		COV	ID-19检	测任务	单/ COV	ID-19 \$	Sample I	Nucleic .	Acid Te	st Recoi	ď		A1	KA0004817
板号/Plate ID.:20200407-1 排单人Arrange Staff: HZB 排单时间/Time:							提取试剂配液编号Extraction Reagent Batch ID: 0318-E1						B1	KA0003576
sample ID	1	2	3	4	5	6	7	8	9	10	н	L.	C1	KA0004837
А	KA0004817	KA0003944	KA0004804	KA0004845									11	KA0004827
A	1 🛛	9 🗆	17 🗆	25 🗆	33 🛛	41 🛛	49 🛛	57 🛛	<mark>65</mark> □	73 🛛	81 🛛	89 🗆	E1	KA0004810
В	KA0003576	KA0003572	KA0004822	KA0004818									F1	KAD. 3578
D	2 🗆	10 🗌	19 🗆	26 🗆	34 🛛	42 🛛	50 🗆	58 🗆	<mark>66</mark> □	74 🛛	82 🛛	90 🗆	G1	KA0004833
с	KA0004837	KA0003585	Blank	KA0003591									H1	KA0003586
C	3 🛛	11 🛛	19 🗆	27 🛛	35 🗆	43 🛛	51 🛛	<u>59</u> 🛛	<mark>67</mark> □	75 🛛	83 🛛	91 🛛	A2	KA0003944
D	KA0004827	KA0003583	KR000+8+5	KA0003210									B2	KA0003572
ע	4 🛛	12 🛛	20 🗆	28 🗆	36 🛛	44 🛛	52 🗆	<u>60</u>	68 🛛	76 🛛	84 🛛	92 🗆	C2	KA0003585
Е	KA0004810	KA0003574	KA0004842										D2	KA0003583
E	5 🗆	13 🛛	21 🗆	29 🗆	37 🛛	45 🛛	53 🛛	<u>61</u>	<mark>69</mark> □	77 🛛	85 🛛	93 🛛	E2	KA0003574
F	KA0003578	KA0004844	KA0004806										F2	KA0004844
г	6 🗌	14 🛛	22 🗆	30 🗆	38 🗆	46 🛛	54 🛛	62 🛛	70 🗆	78 🛛	86 🛛	94 🛛	G2	KA0003597
G	KA0004833	KA0003597	KA0004840										H2	KA0003214
0	7 🛛	15 🛛	23 🗆	31 🗆	39 🗆	47 🛛	55 🗆	63 🛛	71 🛛	79 🛛	87 🛛	95 🗆	A3	KA0004804
Н	KA0003586	KA0003214	KA0004830									PC	B3	KA0004822
н	8 🗆	16 🛛	24 🗆	32 🗆	40 🗆	48 🗆	56 🗆	64 🛛	72 🗆	80 🗆	88 🗆	96 🛛	C3	Blank
备注/Remark: qPCRit济幅浓编号/qPCR Reagant Mix Batch ID: 0318-Q1									D3	KA0004843				
取祥操作人/Sampling Staff. 复核人/Verifier. 取样时间/Sampling Time:									E3	KA0004842				
提取操作人/Nucleic Acid Extraction Staff: 复核人/Verifier: 提取仪器编号/Nucleic Acid Extraction MGISP-960 No.: 6 ト机操作人 aPCR Operator: aPCR(公编号/aPCR Machine No.: 上机时间/aPCR Experiment Time:											F3 G3	KA0004806 KA0004840		

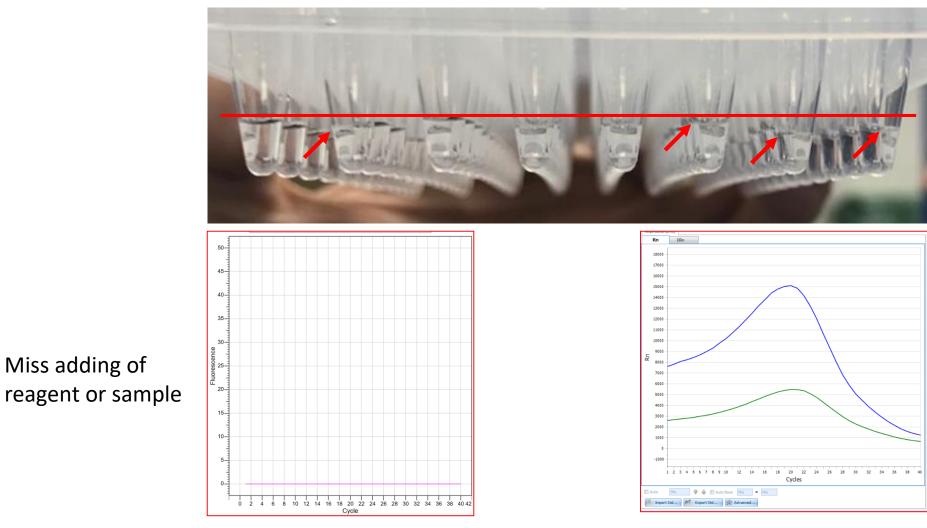


Sample layout for RNA extraction

#### Sample loading verified by two technicians

## Check points - RNA extraction & RT-PCR reaction

#### Sample volume matters!



Evaporation

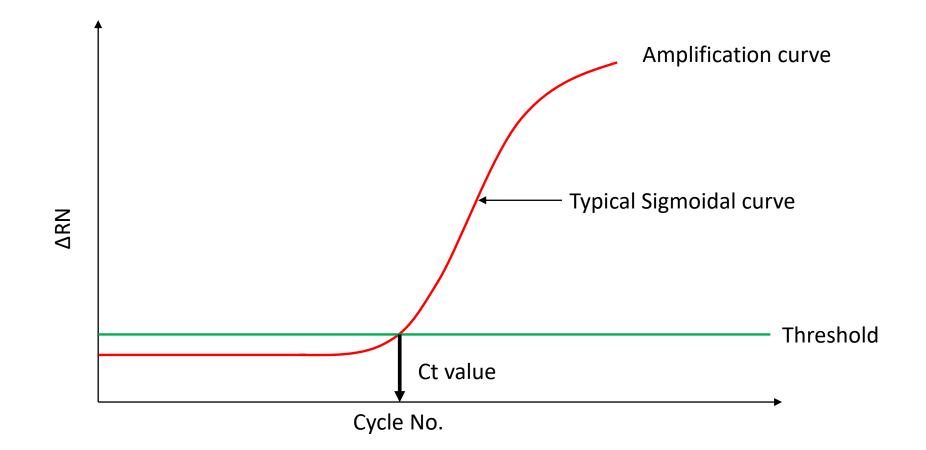
NO Signal Captured

Miss adding of

#### **Unusual Amplification Plot**

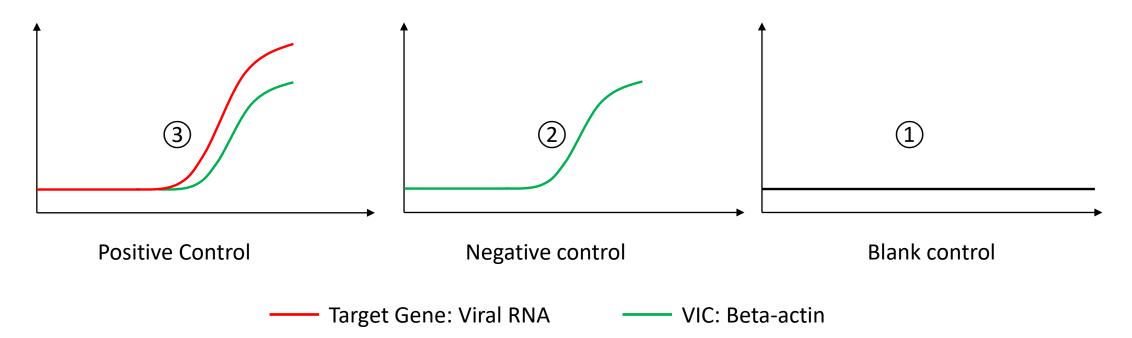
#### Quality control of test data - Key Concepts

- **Baseline**: The signal level during the initial cycles of PCR, usually cycles 3 to 15, in which there is little change in fluorescent signal.
- Threshold: A statistically significant increase over the calculated baseline signal. Usually sets the threshold at 10 times the standard deviation of the fluorescence value of the baseline.
- **Ct**: Cycle number at which the fluorescent signal of the reaction crosses the threshold. So no cross, no Ct.



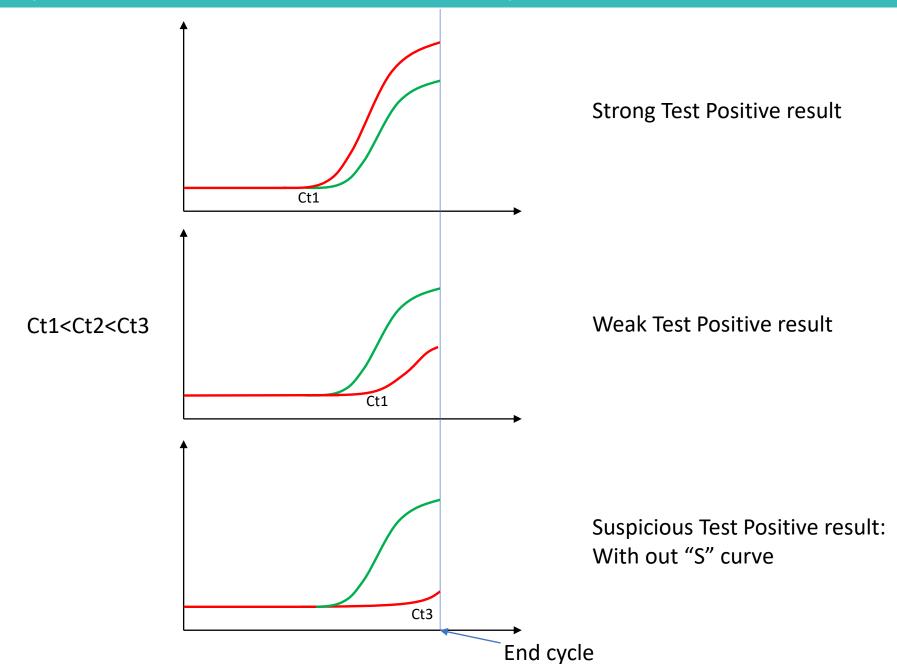
#### Quality control of test data – Control samples

- Positive Control: To check whether Viral RNA can be detected
- Blank control: To monitor the contamination level or background noise level
- Negative control: To check whether beta-actin gene can be amplified successfully without Viral RNA signal

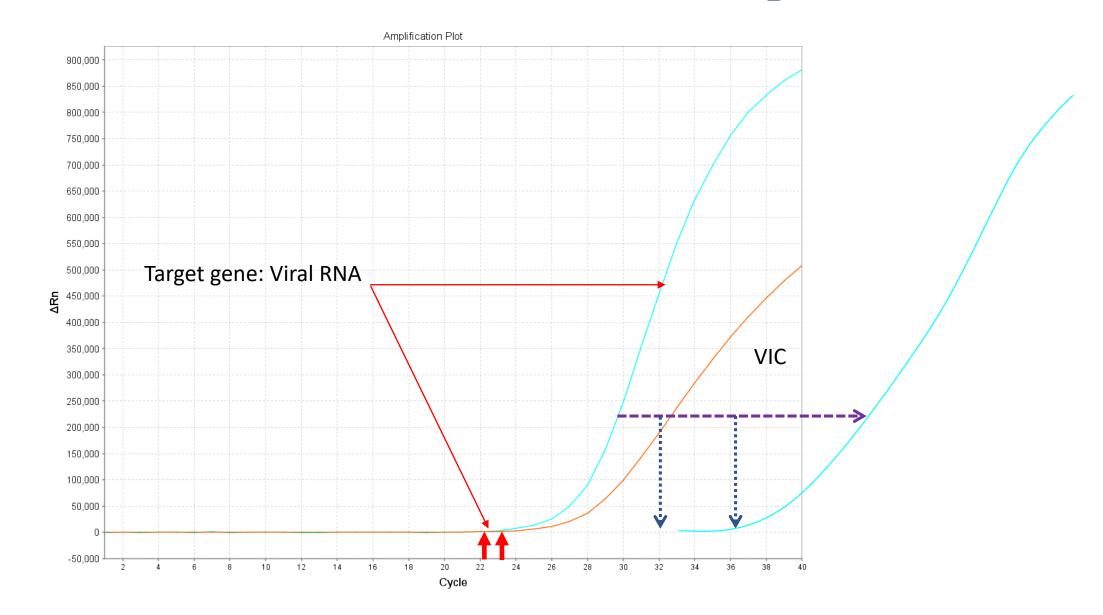


Both viral RNA and beta-actin DNA/RNA should be extracted and then amplified during qPCR reaction, which realizes the quality control of nucleic acid extraction. No beta-actin gene amplification or unusual amplification plot will lead to retest.

#### Quality control of test data – Example of result with Ct value



# Quality control of test data – Example of result with Ct value

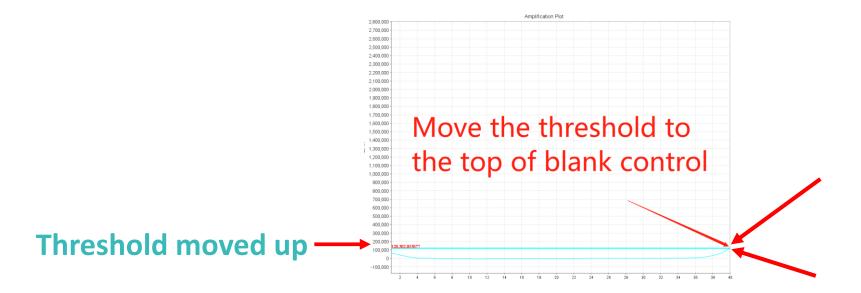


#### Why non-specific amplification in blank control?

- 1) Primer design is not perfect causing primer dimer formation leading to amplification in the end of PCR cycles;
- 2) Dye in the reagent kit partly degrades evidently in the end of cycles;
- 3) Primer in the kit is too concentrated, thus generating non-specific peaks for blank controls;
- 4) Slight contamination during sample mixing;
- 5) Lab environment is not clean enough with accumulated DNA/RNA aerosol;
- 6) 7500 is not well calibrated;

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- Always, \*repeating the test and \* thoroughly clean the laboratory area and related equipment will alleviate this situation. Separating RNA extraction and reagent mixing in different Bio-safety cabinets is suggested.
- Manually move the threshold to cover the noise and reanalyse Ct value for the rest of sample is suggested before data interpretation.



# THANK YOU!